

Expression of Oct4 Protein in Astrocytic Tumours-Histopathologic and Immunohistochemical Study

MENAR M AL-SAYED AYOUB¹, SAMAR ABDEL-MONEIM AL-SHEIKH², LUBNA OMAR AL-FAROUK ABDEL-SALAM³,
ENGY SAMIR MOHAMED ABDEL-MONEIM AL-HARIRY⁴

ABSTRACT

Introduction: The majority of gliomas in adults are resistant to therapy. Therapy resistance has been attributed to the presence of cancer stem cells (CSCs). Oct4 is a transcription factor required for maintaining the pluripotency and self-renewal of CSCs. It was found to be highly expressed in astrocytomas and other tumours.

Aim: Assessment of immunohistochemical expression of Oct4 protein, being a target antigen for cancer immunotherapy of astrocytic tumours, and correlation between Oct4 expression and the grades of astrocytoma.

Materials and Methods: In this retrospective cross sectional study, 66 paraffin embedded biopsies of astrocytomas were collected including; 12 cases of grade I, 19 cases of grade II,

15 cases of grade III and 20 cases of grade IV. All the cases were studied immunohistochemically using anti-Oct4 protein antibody. Chi-square test or Fisher's exact test was used to examine the relation between qualitative variables. All tests were two-tailed. Only a p-value <0.05 was considered significant.

Results: Oct4 was expressed in 30 cases. The higher the grade of astrocytoma, the more was the expression (p-value=0.006) and the percentage of positive tumour cells (p-value=0.039). The expression was both nuclear and cytoplasmic. The intensity of expression was variable within the same tumour. Oct4 was not expressed in normal brain tissue.

Conclusion: Oct4 is a stem cell marker that is involved in the development of CSCs, and accordingly, the development of many tumours including astrocytomas.

Keywords: Astrocytoma, Normal brain tissue, Oct4, Stem cells

INTRODUCTION

Gliomas are the most common primary brain tumours. They were classified by the WHO in 2007 into; astrocytic tumours, oligodendroglial tumours, ependymal tumours and neuronal and mixed neuronal glial tumours [1].

Despite advances in treatment strategies, gliomas remain one of the most fatal and therapy resistant tumours. Resistance to therapy was attributed to the presence of cancer stem cells (CSCs). These cells are immortal, undifferentiated cells similar to early embryonic cells and they have an exclusive ability to self renew and give rise to tumorigenic cancer cells [2]. They are thought to be the determinants for the occurrence, development, recurrence as well as therapy response of gliomas [3, 4].

Oct4, also known as Oct3 or POU5F1, is a member of the POU (Pit, Oct, Unc) transcription factor family encoded by the POU5F1 gene (6p2113) and was first identified in 1990 [5, 6]. The expression of Oct4 is restricted to pluripotent stem cells and is downregulated when differentiation is initiated during embryonic development. It is undetectable in adult normal tissue [7]. Oct4 is regarded as a gatekeeper at early mammalian development [8] and regulates the self-renewal and pluripotency of human embryonic stem cells (ESCs) [9, 10]. It plays an important role in maintaining cellular plasticity and promoting the self-renewal and proliferation ability of stem cells [8].

Oct4-positive cells identified in cancer represent CSCs and account for the maintenance and propagation of tumours [11, 12]. It was found that inhibition of Oct4 expression in glioma-initiating cells resulted in suppression of tumour formation and also potentiated sensitivity to conventional chemotherapy [13]. Therefore, in the current study, we aimed to examine the immunohistochemical expression of Oct4 in different types of astrocytoma and correlated the expression with the grade of astrocytoma.

MATERIALS AND METHODS

This retrospective cross sectional study consisted of 66 archived, formalin fixed, paraffin embedded tissue blocks of astrocytoma (12 as grade I, 19 as grade II, 15 as grade III and 20 as grade IV), collected from pathology department, faculty of medicine, Cairo university, through the period from January 2013 to December 2013. The study was approved by the ethical committee of faculty of medicine, Cairo university.

Each paraffin embedded tissue block was recut by rotatory microtome at 5 microns thickness and was mounted on a glass slide and stained with haematoxylin and eosin (H&E) for routine histopathological examination. The astrocytomas were classified according to 2007 WHO classification [1].

Immunohistochemistry

Paraffin embedded tissue sections were cut at 4 microns thickness and mounted on poly-L Lysine coated slides, and were deparaffinized by fresh xylene, followed by rehydration in graded alcohol. Subsequently, blocking of endogenous peroxidase was done by treating the sections with peroxide block for 15 minutes in room temperature, followed by antigen retrieval for 15 minutes in pressure cooker. Then, the sections were incubated with power blocks for 15 minutes, and primary anti-Oct4 antibody (Ventana, 760-4392), ready to use, was incubated for 40 minutes. DAB chromogen was then incubated for 5-10 minutes. Finally, slides were washed and haematoxylin was used as a counterstain.

High power (x400) was specifically used for examining the intensity of staining which was categorized into (weak, moderate and strong). Expression of Oct4 immunostaining was identified by both nuclear and cytoplasmic staining. 5% was used as the scoring point for calculating the percentage of positive cells. Any number of immune-reactive cells with any intensity of staining was considered positive. The percentage of positive cells was calculated as <5% or

≥5% in both low grade (GII and GIII) and high grade (GIII and GIV) astrocytomas. No other previous papers mentioned about scoring criteria. Section of testicular seminoma was used as a positive control of Oct4.

STATISTICAL ANALYSIS

Statistical analysis was done using IBM® SPSS® Statistics version 22 (IBM® Corp., Armonk, NY, USA). Numerical data were expressed as mean and standard deviation or median and range as appropriate. Qualitative data were expressed as frequency and percentage. Chi-square test or (Fisher's exact test) was used to examine the relation between qualitative variables. All tests were two-tailed. Only a p-value <0.05 was considered significant.

RESULTS

The 66 biopsies of astrocytoma included in this study were classified according to 2007 WHO classification into; 12 cases of grade I astrocytoma (18.2%), 19 cases of grade II astrocytoma (28.8%), 15 cases of grade III astrocytoma (22.7%) and 20 cases of grade IV astrocytoma (30.3%). The different grades were further subclassified into; 8 cases of pilocytic astrocytoma (12.1%) and 4 cases of subependymal giant cell astrocytoma (SEGA) (6.1%) as grade I-14 cases of diffuse astrocytoma (21.2%) and 4 cases of pleomorphic xanthoastrocytoma (6.1%) as well as one case of pilomyxoid astrocytoma (1.5%) as grade II-15 cases of anaplastic astrocytoma (22.7%) as grade III and finally 16 cases of glioblastoma (24.2%) and 4 cases of gliosarcoma (6.1%) as grade IV. Females represented 52% of the study group.

Out of the 66 biopsies of astrocytoma, 36 (54.5%) were negative for Oct4 and 30 (45.5%) showed positive Oct4 expression [Table/Fig-1]. 10 cases (33.3%) showed nuclear expression and 9 cases (30%) showed cytoplasmic expression, while 11 cases (36.7%) showed both nuclear and cytoplasmic expression. The higher the grade of astrocytoma, the higher was the number of immunoreactive cases (p-value=0.006), as well as the percentage of positive cells in each case (p-value=0.039) [Table/Fig-2]. As for the intensity of immunostaining, it was not much affected by the grade (p-value=0.107) [Table/Fig-3].

Grade	Oct4 expression		Total	p-value
	Positive	Negative		
Grade I	1 (1.5%)	11 (16.7%)	12 (18.2%)	0.006
Grade II	7 (10.6%)	12 (18.2%)	19 (28.8%)	
Grade III	8 (12.1%)	7 (10.6%)	15 (22.7%)	
Grade IV	14 (21.2%)	6 (9.1%)	20 (30.3%)	
Total	30 (45.5%)	36 (54.5%)	66 (100%)	

[Table/Fig-1]: Oct4 expression in relation to the grade of astrocytoma (Fisher's exact test).

Grades	Percentage		Total	p-value
	<5%	≥5%		
Grade I and II	7 (23.3%)	1 (3.3%)	8 (26.7%)	0.039
Grade III and IV	9 (30%)	13 (43.3%)	22 (73.3%)	
Total	16 (53.3%)	14 (46.7%)	30 (100%)	

[Table/Fig-2]: Percentage of positive cells in relation to the grade (Fisher's exact test).

Grade	Intensity			Total	p-value
	Weak	Moderate	Strong		
Grade I and II	5 (16.7%)	1 (3.3%)	2 (6.7%)	8 (26.7%)	0.107
Grade III and IV	4 (13.3%)	7 (23.3%)	11 (36.7%)	22 (73.3%)	
Total	9 (30%)	8 (26.7%)	13 (43.3%)	30 (100%)	

[Table/Fig-3]: Intensity of immunostaining in relation to the grade of astrocytoma (Fisher's exact test).

DISCUSSION

Gliomas account for almost 80% of primary brain tumours and most of them show marked therapy resistance. Therefore, studies are done to detect the pathways involved in their development and progression and the possible implication of these pathways in future improvement of therapy lines.

Despite major improvements in knowledge of the biology of gliomas and the molecular and genetic events implicated in the gliomagenesis, little therapeutic progress has been made in the past years and the survival median for glioblastoma patients remains low [14]. Only a small percentage of cells have the potential to recreate the original tumour to its full heterogeneity, and these cells share phenotypic traits with normal stem cells. These cells have been given the name cancer stem cells (CSCs) [14]. Dysregulation of self-renewal of ESCs plays a key role in generation of CSCs [2,15,16].

Oct4 has been implicated in a variety of cellular functions, including maintaining the pluripotency of ESCs and directing their differentiation to particular cell lineages [16, 17]. Ectopic expression of Oct4 in epithelial tissues causes dysplasia by blocking epithelial stem cell differentiation [16, 18]. In several studies, it was suggested that the adult stem cell, expressing the Oct4 gene, was the target cell to start the carcinogenic process [7,11,19].

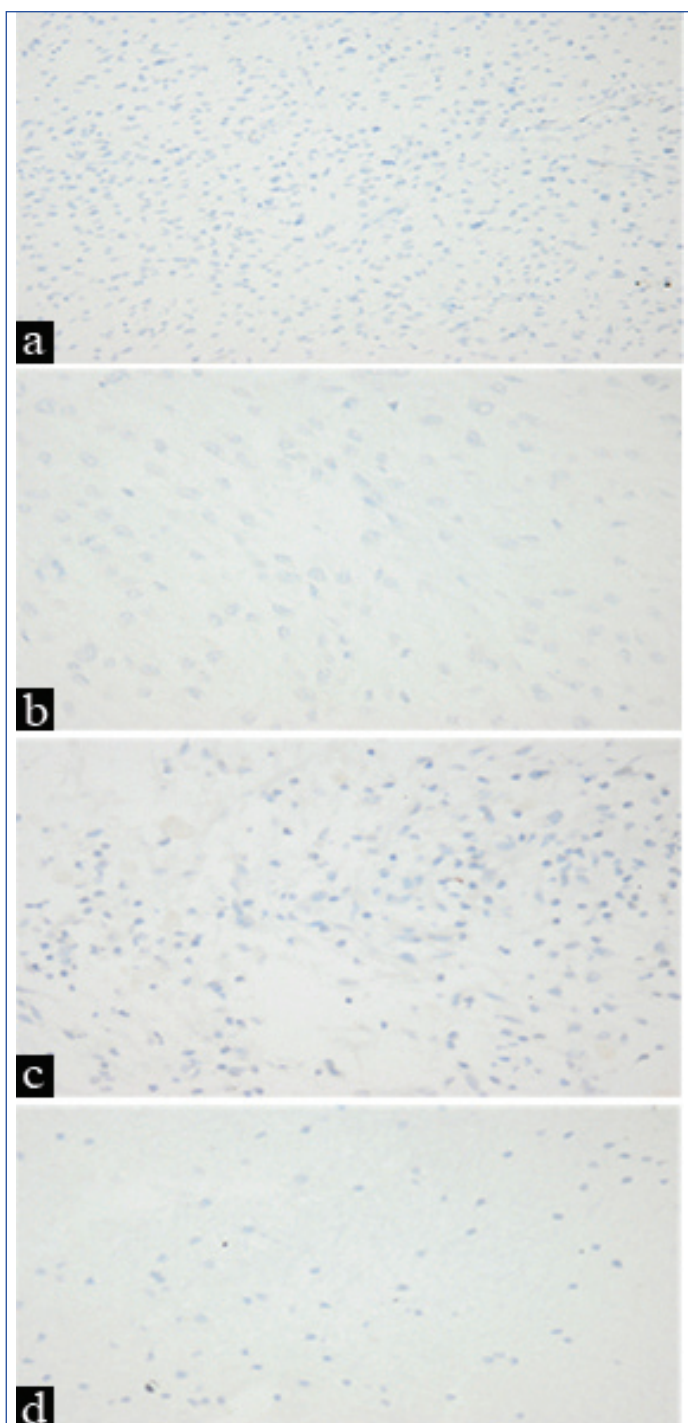
In this study, 66 biopsies of astrocytoma were collected and examined histopathologically, and were classified according to 2007 WHO classification [1] to investigate the immunohistochemical expression of Oct4 protein in different grades of astrocytoma.

The mean age for the patients enrolled in this study was 35±19.4 and it ranged between 2 years and 78 years. Females represented the higher group, as we had 34 females (52%), compared to 32 males (48%) of the study group. Subtypes of astrocytoma of higher grades showed a predilection for higher age group (p-value=0.006), but no specific gender preference (p-value=0.671). No statistical relation was detected between Oct4 expression and age or sex. No other comparative previous studies were available in this regard.

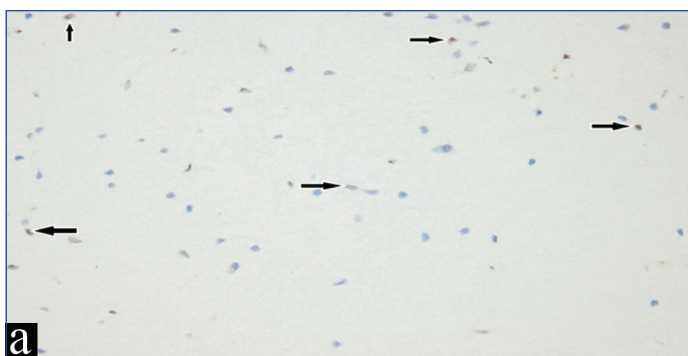
On examination of Oct4 immunohistochemical expression, 36 cases (54.5%) were negative [Table/Fig-4a-d] while 30 cases (45.5%) showed positive expression. The pattern of expression was variable. 10 cases (33.3%) showed nuclear expression [Table/Fig-5a-d], while 9 cases (30%) showed cytoplasmic expression [Table/Fig-6a,b] and 11 cases (36.7%) showed both [Table/Fig-7a-d]. Contrarily, Yuji et al. reported positive expression in 95% of cases, while, coping with the current study, the expression was mainly nuclear, but cytoplasmic expression was also detected [20]. Also, Zhanhui et al. reported expression in all 41 gliomas included in his study and the expression was exclusively nuclear [16]. Oct4 was expressed in all 114 astrocytomas included in the study made by Jeanette et al. They similarly reported that the expression was mainly nuclear, but cytoplasmic expression could be seen too [21]. On the other hand, Oct4 expression was detected only in a few cells in grade IV as reported by Shidou et al. and it was both nuclear and cytoplasmic [22].

Oct4 intensity of staining was found to be higher in higher grades of astrocytoma. Most cases showed moderate and strong intensity [Table/Fig-5d, 6b, 7a-d]. On the other hand, in low grade astrocytomas, the intensity was predominantly weak [Table/Fig-5a,b], while only 3 cases showed higher intensity [Table/Fig-6a]. Similarly, Zhanhui et al., and Jeanette et al., reported similar results [16,21].

The percentage of positive cells was found to be related to the grade with a significant p-value=0.039, which agreed with the results reported by Zhanhui et al., Yuji et al. and Jeanette et al. [16, 20, 21]. On the other hand, Shidou et al., reported that Oct4 expression was only detected in a few cells in grade IV but not detected in any of the lower grades [22].

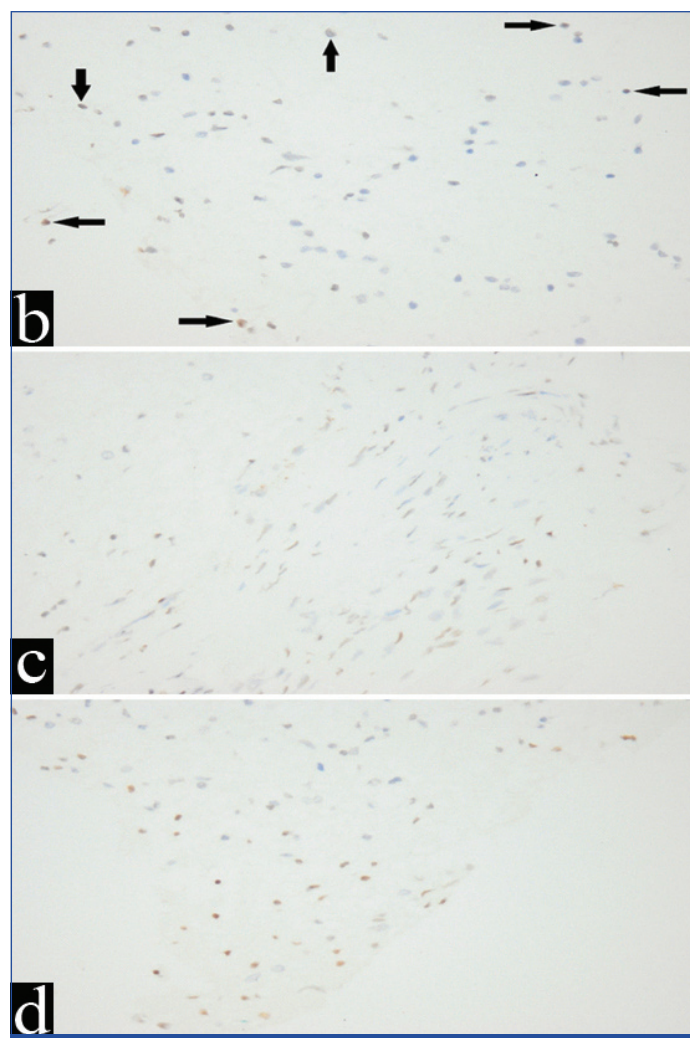


[Table/Fig-4]: a) Pilocytic astrocytoma, negative staining for Oct4 (IHC, x200); b) Subependymal giant cell astrocytoma, negative staining for Oct4 (IHC, x400); c) Pilomyxoid astrocytoma, negative staining for Oct4 (IHC, x400); d) Diffuse astrocytoma, negative staining for Oct4 (IHC, x400).

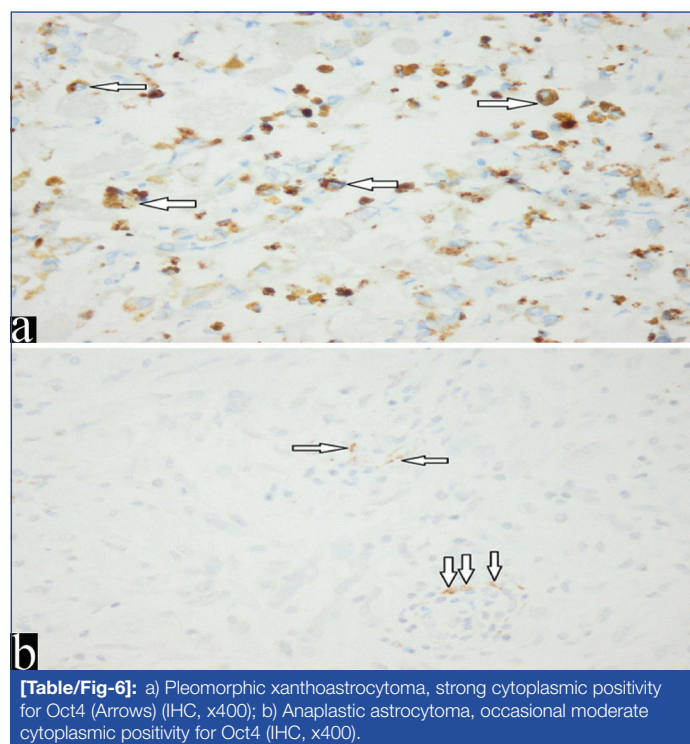


[Table/Fig-5a]: Pilocytic astrocytoma, occasional weakly positive nuclear staining for Oct4 (IHC, x400).

As regard the intensity of Oct4 expression, it was widely variable within the different grades in this study, and even within the same case, but it was not much affected by the grade (p-value=0.107).



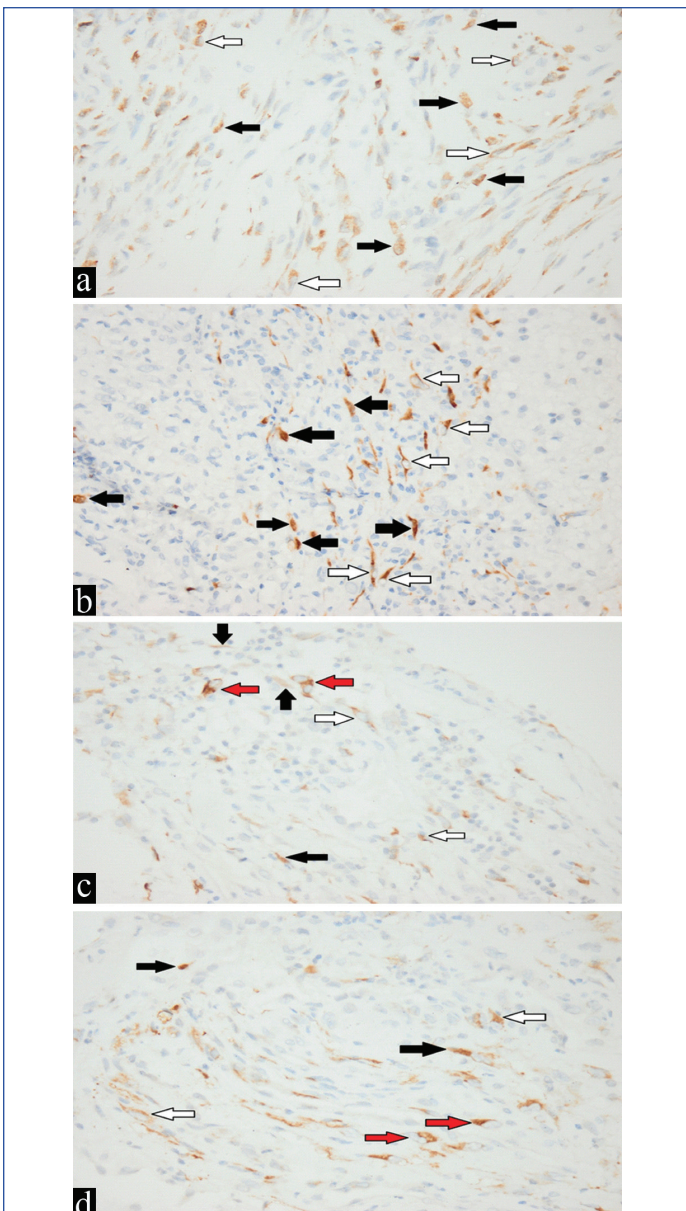
[Table/Fig-5b-d]: b) Diffuse astrocytoma, weakly positive nuclear staining for Oct4 (IHC, x400); c) Anaplastic astrocytoma, weak nuclear staining for Oct4 (IHC, x400); d) Anaplastic astrocytoma, moderate nuclear staining for Oct4 (IHC, x400).



[Table/Fig-6]: a) Pleomorphic xanthoastrocytoma, strong cytoplasmic positivity for Oct4 (Arrows) (IHC, x400); b) Anaplastic astrocytoma, occasional moderate cytoplasmic positivity for Oct4 (IHC, x400).

Jeanette et al. reported similar results, while Zhanhui et al. and Yuji et al. reported that the intensity was much higher in higher grades than in low grades [16, 20, 21].

No expression was detected in normal brain tissue in this study, keeping with Zhanhui et al. findings [16].



[Table/Fig-7a-d]: a) Glioblastoma, moderate nuclear (black arrows) and cytoplasmic (white arrows) staining (IHC, X400) b) Glioblastoma, strong nuclear (black arrows) and cytoplasmic (white arrows) staining (IHC, X400); c) Gliosarcoma, moderate nuclear staining (black arrows) and strong cytoplasmic (red arrows) and moderate cytoplasmic (white arrows) staining (IHC, X400); d) Gliosarcoma, strong nuclear staining (black arrows) and strong cytoplasmic (red arrows) and moderate cytoplasmic (white arrows) staining (IHC, X400).

LIMITATION

The limitation of this study was, not being able to conduct it according to the new molecular classification of astrocytomas in 2016 WHO classification due to limited resources and lack of references in this regard.

CONCLUSION

Oct4 is a transcription factor involved in the pluripotency of embryonic stem cells. Its aberrant expression in adult tissues

is thought to be involved in the development of CSCs and the development of many tumours including astrocytomas. The expression is higher, the higher the grade of astrocytoma. Further studies should be carried out to detect the effect of suppression of Oct4 on the progress of astrocytomas and the possible benefit of immunotherapy.

REFERENCES

- [1] Perry A, Louis DN, Scheithauer BW, Budka H, von Dörmeling A. Meningiomas, in Louis DN, Ohgaki H, Wiestler OD, et al. (eds). World Health Organization Classification of Tumours of the Central Nervous System, ed 4. Lyon: IARC, 2007;17:164.
- [2] Al-Hajj M, Clarke MF. Self-renewal and solid tumor stem cells. *Oncogene*. 2004;23: 274-7282.
- [3] Singh SK, Clarke ID, Terasaki M, Bonn VE, Hawkins C, Squire J, et al. Identification of a cancer stem cell in human brain tumors. *Cancer Res*. 2003;63:5821-5828.
- [4] Singh SK, Clarke ID, Hide T and Dirks PB. Cancer stem cells in nervous system tumors. *Oncogene*. 2004;23:7267-7273.
- [5] Schöler H, Ruppert S, Suzuki N, et al. New type of POU domain in germline-specific protein Oct-4. *Nature*. 1990;344:435-439.
- [6] Jinlong S, Wei S, Lanchun N, Xide X, Xing S, et al. OCT4 is epigenetically regulated by DNA hypomethylation of promoter and exon in primary gliomas. *Oncology Reports*. 2013. 30:201-206.
- [7] Tai MH, Chang CC, Kiupel M, Webster JD, Olson LK, Trosko JE. Oct4 expression in adult human stem cells: Evidence in support of the stem cell theory of carcinogenesis. *Carcinogenesis*. 2005;26:495-502.
- [8] Pesce M and Schöler HR. Oct-4: gatekeeper in the beginnings of mammalian development. *Stem Cells*. 2009;19:271-278.
- [9] Takeda J, Seino S and Bell GT. Human Oct3 gene family: cDNA sequences, alternative splicing, gene organization, chromosomal location, and expression at low levels in adult tissues. *Nucleic Acids Res*. 1992;20:4613-4620.
- [10] Babaie Y, Herwig R, Greber B, et al. Analysis of Oct4-dependent transcriptional networks regulating self-renewal and pluripotency in human embryonic stem cells. *Stem Cells*. 2007;25:500-510.
- [11] Webster JD, Yuzbasiyan-Gurkan V, Trosko JE, Chang CC, Kiupel M. Expression of the embryonic transcription factor Oct4 in canine neoplasms: A potential marker for stem cell subpopulations in neoplasia. *Vet Pathol*. 2007;44:893-900.
- [12] Chiou SH, Yu CC, Huang CY, et al. Positive correlations of Oct-4 and Nanog in oral cancer stem-like cells and high-grade oral squamous cell carcinoma. *Clin Cancer Res*. 2008;14:4085-4095.
- [13] Hiroaki I, Tomoki T, Yasushi I, Masamichi T, Nobuhito S, Keiji M, et al. Glioma-initiating Cells Retain Their Tumorigenicity through Integration of the Sox Axis and Oct4 Protein. *The Journal of Biological Chemistry*. 2011; 286(48):41434-41441.
- [14] Morfouace M, Lallier L, Oliver L, Cheray M, Pecqueur C, Cartron PF et al. Control of glioma cell death and differentiation by PKM2-Oct4 interaction. *Cell Death and Disease*. 2014;5:e1036; doi:10.1038/cddis.561.
- [15] Bar EE, Chaudhry A, Lin A, Fan X, Schreck K, Matsui W, et al. Cyclopamine-mediated hedgehog pathway inhibition depletes stem-like cancer cells in glioblastoma. *Stem Cells*. 2007; 5:2524-2533.
- [16] Zhanhui D, Deyong J, Shangming L, Fuwu W, Gang L, Yanmin Z, et al. Oct4 is Expressed in Human Gliomas and Promotes Colony Formation in Glioma Cells. *GLIA*. 2009;57:724-733.
- [17] Niwa H. How is pluripotency determined and maintained? *Development*. 2007;134:635-646.
- [18] Hochedlinger K, Yamada Y, Beard C, Jaenisch R. Ectopic expression of Oct-4 blocks progenitor-cell differentiation and causes dysplasia in epithelial tissues. *Cell*. 2005;121:465-477.
- [19] Atlasi Y, Mowla SJ, Ziaee SA, Bahrami AR. OCT-4, an embryonic stem cell marker, is highly expressed in bladder cancer. *Int J Cancer*. 2007;120:1598-1602.
- [20] Yuji G, Shangming L, Ping W, Shidou Z, Fuwu W, Lujun B, et al. Expression profile of embryonic stem cell-associated genes Oct4, Sox2 and Nanog in human gliomas. *Histopathology*. 2011;59:763-775.
- [21] Jeanette K P, Per J, Mia D S and Bjarne W K. Expression and Prognostic Value of Oct-4 in Astrocytic Brain Tumors. *PLoS One*. 2016;11(12):e0169129.
- [22] Shidou Z, Qiuhuan Y, Hongbo H, Yuji G, Shangming L, Yanmin Z, et al. Expression of OCT4 pseudogenes in human tumours: lessons from glioma and breast carcinoma. *J Pathol*. 2011;223:672-682.

PARTICULARS OF CONTRIBUTORS:

1. Professor, Department of Pathology, Faculty of Medicine, Cairo University, Cairo, Egypt.
2. Assistant Professor, Department of Pathology, Faculty of Medicine, Cairo University, Cairo, Egypt.
3. Assistant Professor, Department of Pathology, Faculty of Medicine, Cairo University, Cairo, Egypt.
4. PhD Student, Department of Pathology, Faculty of Medicine, Cairo University, Cairo, Egypt.

NAME, ADDRESS, E-MAIL ID OF THE CORRESPONDING AUTHOR:

Engy Samir Mohamed Abdel-Moneim Al-Hariry,
16 Fatma Roshdy St-Al-Haram-Giza-Floor 5, Apartment 603, Cairo, Egypt.
E-mail: drengy2008@hotmail.com

FINANCIAL OR OTHER COMPETING INTERESTS: None.

Date of Submission: **Dec 19, 2017**

Date of Peer Review: **Feb 21, 2018**

Date of Acceptance: **Aug 16, 2018**

Date of Publishing: **Nov 01, 2018**